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=> s (antibody or immunoglobulin) (P) (hydrophilic)

L1 505481 (ANTIBODY OR IMMUNOGLOBULIN) (P) (HYDROPHILICITY OR SOLUBIL? OR
YIELD OR RECOVERY OR EXPRESSION OR INCREAS?)

=> s 11 (P) (domain or fragment)
L2 47308 L1 (P) (DOMAIN OR FRAGMENT)

=> s 12 (P) DNA
L3 7807 L2 (P) DNA

=> s 13 (P) interface
L4 17 L3 (P) INTERFACE

=> dup rem 14
PROCESSING COMPLETED.

PROCESSING COMPLETED FOR L4
L5 6 DUP REM L4 (11 DUPLICATES REMOVED)

=> dis 15 1-6 kwic ibib abs

ANSWER 1 OF 6 CAPLUS COPYRIGHT 2001 ACS
The present invention relates to the modi

the present invention relates to IgSF domains, IgSF fragments and fusion proteins thereof, esp. to the modification of antibody derivs., so as to improve their solv., and hence the yield, and ease of handling. The inventors have found that this can be achieved by making the region which comprises the interface with domains adjoined to said IgSF domain in a larger fragment or a full IgSF protein, and which becomes exposed in the IgSF domain, more hydrophilic by modification. The present invention describes DNA sequences encoding modified IgSF domains or fragments and fusion proteins thereof, vectors and hosts contg. these DNA sequences, IgSF domains or fragments or fusion proteins obtainable by expressing said DNA sequences in suitable expression systems, and a method for modifying IgSF domains, so as to improve their solv., expressibility and ease of handling.

ACCESSION NUMBER: 1998:71159 CAPLUS
DOCUMENT NUMBER: 128:139760
TITLE: Immunoglobulin superfamily domains and fragments with increased solubility
INVENTOR(S): Pluckthun, Andreas; Nieba, Lars; Honegger, Annemarie
PATENT ASSIGNEE(S): Morphosys Gesellschaft Fur Proteinoptimierung M.b.H.,
Germany; Pluckthun, Andreas; Nieba, Lars; Honegger,
Annemarie
SOURCE: PCT Int. Appl., 61 pp.

CODEN: PIXXD2
DOCUMENT TYPE: Patent
LANGUAGE: English
FAMILY ACC. NUM. COUNT: 1
PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9802462	A1	19980122	WO 1997-EP3792	19970716
W: CA, JP, US				
RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
EP 938506	A1	19990901	EP 1997-934467	19970716
R: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,				
IE, FI				
JP 2000516452	T2	20001212	JP 1998-505618	19970716
PRIORITY APPLN. INFO.:			EP 1996-111441	A 19960716
			WO 1997-EP3792	W 19970716

AB The present invention relates to the modification of Ig superfamily (IgSF) domains, IgSF fragments and fusion proteins thereof, esp. to the modification of antibody derivs., so as to improve their solv., and hence the yield, and ease of handling. The inventors have found that this can be achieved by making the region which comprises the interface with domains adjoined to said IgSF domain in a larger fragment or a full IgSF protein, and which becomes exposed in the IgSF domain, more hydrophilic by modification. The present invention describes DNA sequences encoding modified IgSF domains or fragments and fusion proteins thereof, vectors and hosts contg. these DNA sequences, IgSF domains or fragments or fusion proteins obtainable by expressing said DNA sequences in suitable expression systems, and a method for modifying IgSF domains, so as to improve their solv., expressibility and ease of handling.

L5 ANSWER 2 OF 6 MEDLINE DUPLICATE 1
AB . . . beta and transforming growth factor-beta (TGF beta) in regulating TIMP-1, TIMP-3, and 92-kDa type IV collagenase messenger ribonucleic acid (mRNA) expression in human endometrial stromal cells using quantitative competitive PCR. Confluent stromal cell cultures treated with progesterone and estradiol for 9 days were stimulated with IL-1 beta, IL-1 beta plus anti-IL-1 beta antibody, TGF beta, and TGF beta plus anti-TGF beta antibody for an additional 24 h. Competitive complementary DNA fragments were constructed by deletion of a defined fragment from each of the target complementary DNA sequences and coamplified in quantitative competitive PCR as an internal standard. TIMP-1 and TIMP-3, but not 92-kDa type IV collagenase mRNA was only expressed after stimulation with IL-1 beta. IL-1 beta both augmented 92-kDa type IV collagenase mRNA expression and decreased TIMP-1 and TIMP-3 mRNA expression. Conversely, TGF beta augmented TIMP-1 and TIMP-3 mRNA expression, but did not affect 92-kDa type IV collagenase expression. IL-1 and TGF beta-mediated changes were both neutralized by specific antibodies. These results provide indirect evidence that IL-1 and TGF beta may play crucial roles at the embryo-maternal interface during trophoblast invasion by regulating stromal cell expression of TIMP-1, TIMP-3, and 92-kDa type IV collagenase, all of which are known to be important in trophoblast invasion.

ACCESSION NUMBER: 1998251593 MEDLINE
DOCUMENT NUMBER: 98251593 PubMed ID: 9589682

TITLE: Cytokine-mediated regulation of 92-kilodalton type IV collagenase, tissue inhibitor or metalloproteinase-1 (TIMP-1), and TIMP-3 messenger ribonucleic acid expression in human endometrial stromal cells.

AUTHOR: Huang H Y; Wen Y; Irwin J C; Kruessel J S; Soong Y K; Polan M L

CORPORATE SOURCE: Department of Gynecology and Obstetrics, Stanford University Medical Center and School of Medicine, California 94305, USA.

CONTRACT NUMBER: HD-31575 (NICHD)
SOURCE: JOURNAL OF CLINICAL ENDOCRINOLOGY AND METABOLISM, (1998) May; 83 (5) 1721-9.
Journal code: HRB: 0375362. ISSN: 0021-972X.

PUB. COUNTRY: United States
Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English
FILE SEGMENT: Abridged Index Medicus Journals; Priority Journals

ENTRY MONTH: 199806

ENTRY DATE: Entered STN: 19980611
Last Updated on STN: 20000303

Entered Medline: 19980604

AB Interleukin-1 (IL-1) is expressed in human endometrium and has been shown to play an integral role in local cellular interactions during implantation. In addition, the matrix metalloproteinase (MMP) and its inhibitor, the tissue inhibitor of metalloproteinase (TIMP), are crucial during implantation, mediating in vitro trophoblast penetration, and are regulated by several cytokines expressed by trophoblast cells. We have investigated the roles of IL-1 beta and transforming growth factor-beta (TGF beta) in regulating TIMP-1, TIMP-3, and 92-kDa type IV collagenase messenger ribonucleic acid (mRNA) expression in human endometrial stromal cells using quantitative competitive PCR. Confluent stromal cell cultures treated with progesterone and estradiol for 9 days were stimulated with IL-1 beta, IL-1 beta plus anti-IL-1 beta antibody, TGF beta, and TGF beta plus anti-TGF beta antibody for an additional 24 h. Competitive complementary DNA fragments were constructed by deletion of a defined fragment from each of the target complementary DNA sequences and coamplified in quantitative competitive PCR as an internal standard. TIMP-1 and TIMP-3, but not 92-kDa type IV collagenase mRNA, were expressed in stromal cells. The 92-kDa type IV collagenase mRNA was only expressed after stimulation with IL-1 beta. IL-1 beta both augmented 92-kDa type IV collagenase mRNA expression and decreased TIMP-1 and TIMP-3 mRNA expression in a dose-dependent manner. Conversely, TGF beta augmented TIMP-1 and TIMP-3 mRNA expression, but did not affect 92-kDa type IV collagenase expression. IL-1 and TGF beta-mediated changes were both neutralized by specific antibodies. These results provide indirect evidence that IL-1 and TGF beta may play crucial roles at the embryo-maternal interface during trophoblast invasion by regulating stromal cell expression of TIMP-1, TIMP-3, and 92-kDa type IV collagenase, all of which are known to be important in trophoblast invasion.

AB The monoclonal antibody, UB25, recognises a glycoprotein specifically located at the biotrophic interface formed in the *Colletotrichum lindemuthianum*-bean interaction. The antibody labels the walls of intracellular hyphae and the interfacial matrix which separates them from the invaginated host plasma membrane. In . . . are multiples of M(r) 40.5 kDa. A full length cDNA encoding the glycoprotein recognised by UB25 has been isolated by expression cloning and designated CIH1 (*Colletotrichum Intracellular Hypha 1*). In vitro transcription/translation of CIH1, and transfection of mammalian COS cells, showed. . . product in both procedures confirming that the clones isolated were true positives. Southern analysis of bean and *C. lindemuthianum* genomic DNA indicated that the CIH1 glycoprotein is fungally encoded and Northern analysis showed that it is only expressed in planta. Analysis. . . acid sequence of CIH1 indicates the presence of an N-terminal signal sequence and two possible sites for N-glycosylation. The N-terminal domain of the mature protein is rich in proline and contains several short repetitive motifs. CIH1 is thus a fungal proline-rich. . .

ACCESSION NUMBER: 1998388657 MEDLINE
DOCUMENT NUMBER: 98388657 PubMed ID: 9721685
TITLE: Expression cloning of a fungal proline-rich glycoprotein specific to the biotrophic interface formed in the *Colletotrichum*-bean interaction.
AUTHOR: Perfect S E; O'Connell R J; Green E F; Doering-Saad C; Green J R
CORPORATE SOURCE: School of Biological Sciences, University of Birmingham, UK.
SOURCE: PLANT JOURNAL, (1998 July 15) (2) 273-9.
PUB. COUNTRY: ENGLAND: United Kingdom Journal; Article; (JOURNAL ARTICLE)
LANGUAGE: English
FILE SEGMENT: Priority Journals
OTHER SOURCE: GENBANK-AJ001441
ENTRY MONTH: 199810
ENTRY DATE: Entered STN: 19981020 Last Updated on STN: 20000303 Entered Medline: 19981005

AB The monoclonal antibody, UB25, recognises a glycoprotein specifically located at the biotrophic interface formed in the *Colletotrichum lindemuthianum*-bean interaction. The antibody labels the walls of intracellular hyphae and the interfacial matrix which separates them from the invaginated host plasma membrane. In Western blots, UB25 recognises a ladder of bands which are multiples of M(r) 40.5 kDa. A full length cDNA encoding the glycoprotein recognised by UB25 has been isolated by expression cloning and designated CIH1 (*Colletotrichum Intracellular Hypha 1*). In vitro transcription/translation of CIH1, and transfection of mammalian COS cells, showed that UB25 recognized the expressed product in both procedures confirming that the clones isolated were true positives. Southern analysis of bean and *C. lindemuthianum* genomic DNA indicated that the CIH1 glycoprotein is fungally encoded and Northern analysis showed that it is only expressed in planta. Analysis of the deduced amino acid sequence of CIH1 indicates the presence of an N-terminal signal sequence and two possible sites for N-glycosylation. The N-terminal domain of the mature protein is rich in proline and contains several short repetitive motifs. CIH1 is thus a fungal proline-rich glycoprotein which appears to form a cross-linked structure in planta and, as such, resembles plant cell wall proline- and hydroxyproline-rich proteins. Possible functions for the CIH1 protein in the establishment and maintenance of biotrophy are discussed.

LS ANSWER 4 OF 6 MEDLINE DUPLICATE 3
AB . . . ER during fetal uterine development. The purpose of this study was to investigate the pattern of ER gene and protein expression in the human fetal uterus. Uteri were obtained from abortuses (n = 43; range, 10-24 weeks gestation) of women undergoing. . . cultured fetal uterine stroma-like cells (n = 15). Immunolocalization studies were performed on frozen uterine sections using the H222 monoclonal antibody (Abbott Laboratories) directed against the ER (n = 20). Western blotting was used to confirm the identity of the ER. . . = 3. Ligand binding studies were performed using radiolabeled [³H]estradiol (n = 5). Using reverse transcription-polymerase chain reaction, a 263-basepair DNA fragment corresponding to the ER was consistently present in uteri after 15 weeks gestation. By immunohistochemistry, the ER is expressed within the uterine mesenchyme in a discrete cylindrical pattern at the interface of the differentiating endometrial stroma and myometrium. Western blotting confirmed the presence of a protein of an apparent mol wt. . .

ACCESSION NUMBER: 95189894 MEDLINE
DOCUMENT NUMBER: 95189894 PubMed ID: 7883857
TITLE: Ontogeny of the estrogen receptor in the human fetal uterus.
AUTHOR: Glatstein I Z; Yeh J
CORPORATE SOURCE: Department of Obstetrics, Gynecology, and Reproductive Biology, Brigham and Women's Hospital, Boston, Massachusetts 02115.
SOURCE: JOURNAL OF CLINICAL ENDOCRINOLOGY AND METABOLISM, (1995 Mar) 80 (3) 958-64.
PUB. COUNTRY: United States Journal; Article; (JOURNAL ARTICLE)
LANGUAGE: English
FILE SEGMENT: Abridged Index Medicus Journals; Priority Journals
ENTRY MONTH: 199504
ENTRY DATE: Entered STN: 19950425 Last Updated on STN: 19950425 Entered Medline: 19950411

AB The role of estrogen and its receptor in the development of the human fetal reproductive tract is unknown, but it may be involved in uterine maturation. In mouse and guinea pig uteri, studies have identified estrogen receptor (ER) protein during the fetal period. In the human, there are no published data regarding the ER during fetal uterine development. The purpose of this study was to investigate the pattern of ER gene and protein expression in the human fetal uterus. Uteri were obtained from abortuses (n = 43; range, 10-24 weeks gestation) of women undergoing elective termination of pregnancy. Reverse transcription-polymerase chain reaction was performed with whole uteri as well as cultured fetal uterine stroma-like cells (n = 15). Immunolocalization studies were performed on frozen uterine sections using the H222 monoclonal antibody (Abbott Laboratories) directed against the ER (n = 20). Western blotting was used to confirm the identity of the ER protein (n = 3). Ligand binding studies were performed using radiolabeled [³H]estradiol (n = 5). Using reverse transcription-polymerase

chain reaction, a 263-basepair DNA fragment corresponding to the ER was consistently present in uteri after 15 weeks gestation. By immunohistochemistry, the ER is expressed within the uterine mesenchyme in a discrete cylindrical pattern at the interface of the differentiating endometrial stroma and myometrium. Western blotting confirmed the presence of a protein of an apparent mol wt of 66 kilodaltons, the predicted size of the ER. Ligand binding studies for the ER gave a value less than 8 fmol/mg protein. In summary, the ER gene is expressed in the uterus beginning in the early second trimester during fetal development. ER protein is localized in a discrete cylindrical pattern within the developing uterus. The highly specific location of the ER protein together with the messenger ribonucleic acid data suggest a role for the ER in differentiation of the primitive uterine mesenchyme into stromal and myometrial compartments.

L5 ANSWER 5 OF 6 MEDLINE DUPLICATE 4
AB . . . adhesion molecule-1 (ELAM-1), a cell surface glycoprotein expressed by cytokine-activated endothelium, mediates the adhesion of blood neutrophils. A full-length complementary DNA (cDNA) for ELAM-1 has now been isolated by transient expression in COS cells. Cells transfected with the ELAM-1 clone express a surface structure recognized by two ELAM-1 specific monoclonal antibodies (H4/18 and H18/7) and support the adhesion of isolated human neutrophils and the promyelocytic cell line HL-60. Expression of ELAM-1 transcripts in cultured human endothelial cells is induced by cytokines, reaching a maximum at 2 to 4 hours and decaying by 24 hours; cell surface expression of ELAM-1 protein parallels that of the mRNA. The primary sequence of ELAM-1 predicts an amino-terminal lectin-like domain, an EGF domain, and six tandem repetitive motifs (about 60 amino acids each) related to those found in complement regulatory proteins. A similar domain structure is also found in the MEL-14 lymphocyte cell surface homing receptor, and in granule-membrane protein 140, a membrane glycoprotein. . . of a nascent gene family of cell surface molecules involved in the regulation of inflammatory and immunological events at the interface of vessel wall and blood.

ACCESSION NUMBER: 89162047 MEDLINE
DOCUMENT NUMBER: 89162047 PubMed ID: 2466335
TITLE: Endothelial leukocyte adhesion molecule 1: an inducible receptor for neutrophils related to complement regulatory proteins and lectins.
AUTHOR: Bevilacqua M P; Stengelin S; Gimbrone M A Jr; Seed B
CORPORATE SOURCE: Department of Pathology, Brigham and Women's Hospital, Boston, MA 02115.
CONTRACT NUMBER: P01 HL-36028 (NHLBI)
SOURCE: SCIENCE, (1989 Mar 3) 243 (4895) 1160-5.
Journal code: UJ7; 0404511. ISSN: 0036-8075.
PUB. COUNTRY: United States
LANGUAGE: English
FILE SEGMENT: Priority Journals
OTHER SOURCE: GENBANK-M24736
ENTRY MONTH: 198904
ENTRY DATE: Entered STN: 19900306
Last Updated on STN: 19970203
Entered Medline: 19890407

AB Focal adhesion of leukocytes to the blood vessel lining is a key step in inflammation and certain vascular disease processes. Endothelial leukocyte adhesion molecule-1 (ELAM-1), a cell surface glycoprotein expressed by cytokine-activated endothelium, mediates the adhesion of blood neutrophils. A full-length complementary DNA (cDNA) for ELAM-1 has now been isolated by transient expression in COS cells. Cells transfected with the ELAM-1 clone express a surface structure recognized by two ELAM-1 specific monoclonal antibodies (H4/18 and H18/7) and support the adhesion of isolated human neutrophils and the promyelocytic cell line HL-60. Expression of ELAM-1 transcripts in cultured human endothelial cells is induced by cytokines, reaching a maximum at 2 to 4 hours and decaying by 24 hours; cell surface expression of ELAM-1 protein parallels that of the mRNA. The primary sequence of ELAM-1 predicts an amino-terminal lectin-like domain, an EGF domain, and six tandem repetitive motifs (about 60 amino acids each) related to those found in complement regulatory proteins. A similar domain structure is also found in the MEL-14 lymphocyte cell surface homing receptor, and in granule-membrane protein 140, a membrane glycoprotein of platelet and endothelial secretory granules that can be rapidly mobilized (less than 5 minutes) to the cell surface by thrombin and other stimuli. Thus, ELAM-1 may be a member of a nascent gene family of cell surface molecules involved in the regulation of inflammatory and immunological events at the interface of vessel wall and blood.

L5 ANSWER 6 OF 6 MEDLINE DUPLICATE 5
AB . . . The starting model was the refined 2.7 Å structure of unliganded Fab from an autoantibody (BV04-01) with specificity for single-stranded DNA. In the 4-4-20 complex fluorescein fits tightly into a relatively deep slot formed by a network of tryptophan and tyrosine. . . chains. The planar xanthonyl ring of the hapten is accommodated at the bottom of the slot while the phenylcarboxyl group interfaces with solvent. Tyrosine 37 (light chain) and tryptophan 33 (heavy chain) flank the xanthonyl group and tryptophan 101 (light chain). . . haptene-dianion. Formation of an enol-arginine ion pair in a region of low dielectric constant may account for an incremental increase in affinity of 2-3 orders of magnitude in the 4-4-20 molecule relative to other members of an idiotypic family of monoclonal antifluoresceyl antibodies. The phenyl carboxyl group of fluorescein appears to be hydrogen-bonded to the phenolic hydroxyl group of tyrosine 37 of the light chain. A molecule of 2-methyl-2,4-pentanediol (MPD), trapped in the interface of the variable domains just below the fluorescein binding site, may be partly responsible for the decrease in affinity for the hapten in MPD.

ACCESSION NUMBER: 90017460 MEDLINE
DOCUMENT NUMBER: 90017460 PubMed ID: 2508085
TITLE: Three-dimensional structure of a fluorescein-Fab complex crystallized in 2-methyl-2,4-pentanediol.
AUTHOR: Herron J N; He X M; Mason M L; Voss E W Jr; Edmundson A B
CORPORATE SOURCE: Department of Biology, University of Utah, Salt Lake City 84112.
CONTRACT NUMBER: AI 20960 (NIAID)
AI 22898 (NIAID)
CA 19616 (NCI)
+
SOURCE: PROTEINS, (1989) 5 (4) 271-80.
Journal code: PTS; 8700181. ISSN: 0887-3585.

PUB. COUNTRY: United States
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 198911
ENTRY DATE: Entered STN: 19900328
Last Updated on STN: 19980206
Entered Medline: 19891108

AB The crystal structure of a fluorescein-Fab (4-4-20) complex was determined at 2.7 Å resolution by molecular replacement methods. The starting model was the refined 2.7 Å structure of unliganded Fab from an autoantibody (BV04-01) with specificity for single-stranded DNA. In the 4-4-20 complex fluorescein fits tightly into a relatively deep slot formed by a network of tryptophan and tyrosine side chains. The planar xanthonyl ring of the hapten is accommodated at the bottom of the slot while the phenylcarboxyl group interfaces with solvent. Tyrosine 37 (light chain) and tryptophan 33 (heavy chain) flank the xanthonyl group and tryptophan 101 (light chain) provides the floor of the combining site. Tyrosine 103 (heavy chain) is situated near the phenyl ring of the hapten and tyrosine 102 (heavy chain) forms part of the boundary of the slot. Histidine 31 and arginine 39 of the light chain are located in positions adjacent to the two enolic groups at opposite ends of the xanthonyl ring, and thus account for neutralization of one of two negative charges in the haptenic dianion. Formation of an enol-arginine ion pair in a region of low dielectric constant may account for an incremental increase in affinity of 2-3 orders of magnitude in the 4-4-20 molecule relative to other members of an idiotypic family of monoclonal antifluorescein antibodies. The phenyl carboxyl group of fluorescein appears to be hydrogen bonded to the phenolic hydroxyl group of tyrosine 37 of the light chain. A molecule of 2-methyl-2,4-pentanediol (MPD), trapped in the interface of the variable domains just below the fluorescein binding site, may be partly responsible for the decrease in affinity for the hapten in MPD.

=> s Pluckthun A?au or Nieba L?au or Honegger A?au
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L6 651 PLUCKTHUN A?/AU OR NIEBA L?/AU OR HONEGGER A?/AU

=> s 16 and antibod?
L7 313 L6 AND ANTIBOD?

=> s 17 and hydro?
L8 46 L7 AND HYDRO?

=> dup rem 18
PROCESSING COMPLETED FOR L8
L9 20 DUP REM L8 (26 DUPLICATES REMOVED)

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=> dis 19 1-20 ibib abs

L9 ANSWER 1 OF 20 MEDLINE
ACCESSION NUMBER: 2001150051 MEDLINE
DOCUMENT NUMBER: 21103695 PubMed ID: 11162118
TITLE: The scFv fragment of the antibody hu4D5-8: evidence for early premature domain interaction in refolding.
AUTHOR: Jager M; Gehrig P; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut, Universität Zurich,
Winterthurerstr. 190, CH-8057, Zurich, Switzerland.
SOURCE: JOURNAL OF MOLECULAR BIOLOGY, (2001 Feb 9) 305 (5) 1111-29.
Journal code: J6V; 2985088R. ISSN: 0022-2836.
PUB. COUNTRY: England: United Kingdom
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 200103
ENTRY DATE: Entered STN: 20010404
Last Updated on STN: 20010404
Entered PubMed: 20010222
Entered Medline: 20010315

AB Fluorescence spectroscopy and 1H/2H-exchange techniques have been applied to characterize the folding of an scFv fragment, derived from the humanized anti-HER2 antibody hu4D5-8. A stable intermediate, consisting of a native VL domain and an unfolded VH domain, is populated under equilibrium unfolding conditions. A partially structured intermediate, with 1H/2H-exchange protection significantly less than that of the two isolated domains together, is detectable upon refolding the equilibrium-denatured scFv fragment. This means that the domains in the heterodimer do not fold independently. Rather, they associate prematurely before full 1H/2H-exchange protection can be gained. The formation of the native heterodimer from the non-native intermediate is a slow, cooperative process, which is rate-limited by proline cis/trans-isomerization. Unproductive domain association is also detectable after short-term denaturation, i.e. with the proline residues in native conformation. Only a fraction of the short-term denatured protein folds into the native protein in a fast, proline-independent reaction, because of spontaneous proline cis/trans-reisomerization in the early non-native intermediate. The comparison with the previously studied antibody McPC603 has now allowed us to delineate similarities in the refolding pathway of scFv fragments.

L9 ANSWER 2 OF 20 MEDLINE DUPLICATE 1
ACCESSION NUMBER: 2000214282 MEDLINE
DOCUMENT NUMBER: 20214282 PubMed ID: 10752617
TITLE: Direct evidence by H/D exchange and ESI-MS for transient unproductive domain interaction in the refolding of an antibody scFv fragment.

AUTHOR: Jager M; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut, Universitit Zurich, Switzerland.
SOURCE: PROTEIN SCIENCE, (2000 Mar) 9 (3) 552-63.
Journal code: BNW; 9211750. ISSN: 0961-8368.
PUB. COUNTRY: United States
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 200005
ENTRY DATE: Entered STN: 20000606
Last Updated on STN: 20000606
Entered Medline: 20000519

AB The refolding kinetics of a single-chain Fv (scFv) fragment, derived from a stabilized mutant of the phosphorylcholine binding antibody McPC603, was investigated by H/D exchange and ESI-MS and compared with the folding kinetics of its constituting domains V(H) and V(L). Both V(H) and V(L) adopt essentially native-like exchange protection within the dead time of the manual-mixing H/D exchange experiment (10 s) and in the case of V(L), which contains two cis-prolines in the native conformation, this fast protection is independent of proline cis/trans isomerization. At the earliest time point resolvable by manual mixing, fewer deuterons are protected in the scFv fragment than in the two isolated domains together, despite the fact that the scFv fragment is significantly more stable than V(L) and V(H). Full H/D exchange protection in the scFv fragment is gained on a time scale of minutes. This means that the domains in the scFv fragment do not refold independently. Rather, they associate prematurely and in nonnative form, a kinetic trap. Unproductive domain association is observed both after equilibrium- and short-term denaturation. For the equilibrium-denatured scFv fragment, whose native structure formation is dependent on a cis conformation of an interface proline in V(L), this cis/trans isomerization reaction proceeds about one order in magnitude more slowly than the escape from the trap to a conformation where full H/D exchange protection is already achieved. We interpret these data in terms of a general kinetic scheme involving intermediates with and without domain association.

L9 ANSWER 3 OF 20 MEDLINE DUPLICATE 2
ACCESSION NUMBER: 1999457490 MEDLINE
DOCUMENT NUMBER: 99457490 PubMed ID: 10525411
TITLE: Insight into odorant perception: the crystal structure and binding characteristics of antibody fragments directed against the musk odorant traseolide.
AUTHOR: Langedijk A C; Spinelli S; Anguille C; Hermans P; Nederlof J; Butenandt J; Honegger A; Cambillau C; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut, Universitat Zurich, Winterthurerstrasse 190, Zurich, CH-8057, Switzerland.
SOURCE: JOURNAL OF MOLECULAR BIOLOGY, (1999 Oct 14) 292 (4) 855-69.
Journal code: J6V; 2985088R. ISSN: 0022-2836.
PUB. COUNTRY: ENGLAND: United Kingdom
LANGUAGE: English
FILE SEGMENT: Priority Journals
OTHER SOURCE: PDB-1C12
ENTRY MONTH: 199911
ENTRY DATE: Entered STN: 20000111
Last Updated on STN: 20000111
Entered Medline: 19991104

AB Monoclonal antibodies were elicited against the small hydrophobic hapten traseolide, a commercially available musk fragrance. Antibody variable region sequences were found to belong to different sequence groups, and the binding characteristics of the corresponding antibody fragments were investigated. The antibodies M02/01/01 and M02/05/01 are highly homologous and differ in the binding pocket only at position H93. M02/05/01 (H93 Val) binds the hapten traseolide about 75-fold better than M02/01/01 (H93 Ala). A traseolide analog, missing only one methyl group, does not have the characteristic musk odorant fragrance. The antibody M02/05/01 binds this hapten analog about tenfold less tightly than the original traseolide hapten, and mimics the odorant receptor in this respect, while the antibody M02/01/01 does not distinguish between the analog and traseolide. To elucidate the structural basis for the fine specificity of binding, we determined the crystal structure of the Fab fragment of M02/05/01 complexed with the hapten at 2.6 Å resolution. The crystal structure showed that only van der Waals interactions are involved in binding. The somatic Ala H93 Val mutation in M02/05/01 fills up an empty cavity in the binding pocket. This leads to an increase in binding energy and to the ability to discriminate between the hapten traseolide and its derivatives. The structural understanding of odorant specificity in an antibody gives insight in the physical principles on how specificity for such hydrophobic molecules may be achieved.

L9 ANSWER 4 OF 20 MEDLINE DUPLICATE 3
ACCESSION NUMBER: 1999321993 MEDLINE
DOCUMENT NUMBER: 99321993 PubMed ID: 10390351
TITLE: Removal of the conserved disulfide bridges from the scFv fragment of an antibody: effects on folding kinetics and aggregation.
AUTHOR: Ramm K; Gehrig P; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut, Universitat Zurich, Winterthurerstr. 190, Zurich, CH-8057, Switzerland.
SOURCE: JOURNAL OF MOLECULAR BIOLOGY, (1999 Jan 9) 290 (2) 535-46.
Journal code: J6V; 2985088R. ISSN: 0022-2836.
PUB. COUNTRY: ENGLAND: United Kingdom
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199907
ENTRY DATE: Entered STN: 19990816
Last Updated on STN: 19990816
Entered Medline: 19990730

AB Fluorescence measurements and H/D exchange experiments monitored by mass spectrometry have been applied to investigate the influence of the conserved disulfide bridges on the folding behavior and in vitro aggregation properties of the scFv fragment of the antibody hu4D5-8. A set of four proteins, carrying none, one, or both of the disulfide bridges have been compared regarding their stabilities, folding kinetics and tendency to aggregate. The results show that refolding of all four scFvs is ultimately limited by a slow proline isomerization in the VLdomain, since the native cis -conformation of proline 195 seems to be a prerequisite for formation of the native interface. Starting from short-term denatured protein, with the proline residues in their native conformation, a kinetically trapped intermediate is populated depending on

the conditions, whose rate of conversion is slower than that of the fast-folding molecules. According to deuteron protection patterns determined by mass spectrometry, those domains retaining the disulfide bridge are able to form stable native-like structure, independent of native interface formation. The disulfide-free domains, in contrast, require the native interface for sufficient stabilization. The resistance of the scFvs towards aggregation seems to be critically dependent on the presence of the disulfide bridge in the VHdomain, and thus on the ability of the VHdomain to form stable structure prior to interaction with the VLdomain. The presence of stable VLdomain in combination with a disulfide-free VHdomain appears to further promote aggregation, indicating the involvement of structured domains in the aggregates.

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L9 ANSWER 5 OF 20 CAPLUS COPYRIGHT 2001 ACS
ACCESSION NUMBER: 1999:449739 CAPLUS
DOCUMENT NUMBER: 132:90223
TITLE: SPM for functional identification of individual biomolecules
AUTHOR(S): Ros, Robert; Schwesinger, Falk; Padeste, Celestino;
Pluckthun, Andreas; Anselmetti, Dario;
Guentherodt, Hans-Joachim; Tiefenauer, Louis
CORPORATE SOURCE: Molecular Nanotechnology, Paul Scherrer Institute,
Villigen, Switz.
SOURCE: Proc. SPIE-Int. Soc. Opt. Eng. (1999), 3607 (Scanning and Force Microscopies for Biomedical Applications), 84-89
PUBLISHER: SPIE-The International Society for Optical Engineering
DOCUMENT TYPE: Journal
LANGUAGE: English
AB The identification of specific binding mols. is of increasing interest in the context of drug development based on combinatorial libraries. Scanning Probe Microscopy (SPM) is the method of choice to image and probe individual biomols. on a surface. Functional identification of biomols. is a first step towards screening on a single mol. level. As a model system we use recombinant single-chain Fv fragment (scFv) antibody mols. directed against the antigen fluorescein. The scFv's are covalently immobilized on a flat gold surface via the C-terminal cysteine, resulting in a high accessibility of the binding site. The antigen is immobilized covalently via a long hydrophilic spacer to the silicon nitride SPM-tip. This arrangement allows a direct measurement of binding forces. Thus, closely related antibody mols. differing in only one amino acid at their binding site could be distinguished. A novel SPM-software has been developed which combines imaging, force spectroscopic modes, and online anal. This is a major prerequisite for future screening methods.
REFERENCE COUNT: 18
REFERENCE(S): (1) Allen, S; Biochemistry 1997, V36, P7457 CAPLUS
(2) Anselmetti, D; J Vac Sci Technol B 1994, V12, P1500 CAPLUS
(4) Chilkoti, A; Biophys J 1995, V69, P2125 CAPLUS
(5) Dammer, U; Biophys J 1996, V70, P2437 CAPLUS
(7) Florin, E; Science 1994, V264, P415 CAPLUS
ALL CITATIONS AVAILABLE IN THE RE FORMAT

L9 ANSWER 6 OF 20 CAPLUS COPYRIGHT 2001 ACS
ACCESSION NUMBER: 1998:71159 CAPLUS
DOCUMENT NUMBER: 128:139760
TITLE: Immunoglobulin superfamily domains and fragments with increased solubility
INVENTOR(S): Pluckthun, Andreas; Nieba, Lars;
Honegger, Annemarie
PATENT ASSIGNEE(S): Morphosys Gesellschaft Fur Proteinoptimierung M.b.H.,
Germany; Pluckthun, Andreas; Nieba, Lars; Honegger,
Annemarie
SOURCE: PCT Int. Appl., 61 pp.
DOCUMENT TYPE: Patent
LANGUAGE: English
FAMILY ACC. NUM. COUNT: 1
PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9802462	A1	19980122	WO 1997-EP3792	19970716
W: CA, JP, US RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
EP 938506	A1	19990901	EP 1997-934467	19970716
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
JP 2000516452	T2	20001212	JP 1998-505618	19970716
PRIORITY APPLN. INFO.:			EP 1996-111441	A 19960716
			WO 1997-EP3792	W 19970716

AB The present invention relates to the modification of Ig superfamily (IgSF) domains, IgSF fragments and fusion proteins thereof, esp. to the modification of antibody derivs., so as to improve their solv., and hence the yield, and ease of handling. The inventors have found that this can be achieved by making the region which comprises the interface with domains adjoined to said IgSF domain in a larger fragment or a full IgSF protein, and which becomes exposed in the IgSF domain, more hydrophilic by modification. The present invention describes DNA sequences encoding modified IgSF domains or fragments and fusion proteins thereof, vectors and hosts contg. these DNA sequences, IgSF domains or fragments or fusion proteins obtainable by expressing said DNA sequences in suitable expression systems, and a method for modifying IgSF domains, so as to improve their solv., expressibility and ease of handling.

L9 ANSWER 7 OF 20 MEDLINE
ACCESSION NUMBER: 1998422265 MEDLINE
DOCUMENT NUMBER: 98422265 PubMed ID: 9748318
TITLE: Mutual stabilization of VL and VH in single-chain antibody fragments, investigated with mutants engineered for stability.
AUTHOR: Worn A; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut, Universitat Zurich, Switzerland.
SOURCE: BIOCHEMISTRY, (1998 Sep 22) 37 (38) 13120-7.
Journal code: AOG; 0370623. ISSN: 0006-2960.
PUB. COUNTRY: United States
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199810
ENTRY DATE: Entered STN: 19981029

Last Updated on STN: 2000
Entered Medline: 19981022

AB A set of six mutants of the levan binding single-chain Fv (scFv) fragment A48 (ABPC48), which have the identical light chain but differ gradually in the stability of the heavy chain, was generated. This was achieved by introducing one or both of the stabilizing mutations H-K66R and H-N52S into the VH domain of the A48 wild-type protein, which is naturally missing the conserved disulfide bridge in VH, and into the cysteine-restored variant A48cys scFv. The stabilizing effects of these two mutations in VH, which had been selected in the context of a disulfide-free derivative of this scFv fragment [Proba, K., et al. (1998) J. Mol. Biol. 275, 245-253], were found to be additive and transferable to the cysteine-restored variant of the A48 scFv, thereby generating extremely stable VH domains. The equilibrium denaturation of these scFv fragments was compared with the corresponding isolated VL domain and two of the different isolated VH domains. In the scFv fragment, the VL domain was found to be stabilized by a more stable VH domain, and, conversely, the VH domain was stabilized by a more stable VL domain. A folding intermediate with native-like VH and denatured VL was found at equilibrium, if VH was significantly more stable than VL. In all other cases, a cooperative unfolding of the scFv was observed. We explain this observation with different contributions of intrinsic domain stability and extrinsic stabilization provided by the partner domain in the single-chain antibodies.

L9 ANSWER 8 OF 20 MEDLINE DUPLICATE 4

ACCESSION NUMBER: 1998409447 MEDLINE
DOCUMENT NUMBER: 98409447 PubMed ID: 9737871
TITLE: Factors influencing the dimer to monomer transition of an antibody single-chain Fv fragment.
AUTHOR: Arndt K M; Müller K M; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut, Universität Zurich, Switzerland.
SOURCE: BIOCHEMISTRY, (1998 Sep 15) 37 (37) 12918-26.
Journal code: A061 0370623. ISSN: 0006-2960.
PUB. COUNTRY: United States
Journal; Article; (JOURNAL ARTICLE)
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199810
ENTRY DATE: Entered STN: 19981029
Last Updated on STN: 19981029
Entered Medline: 19981020

AB Antibody single-chain Fv (scFv) fragments are able to form dimers under certain conditions, and the extent of dimerization appears to depend on linker length, antibody sequence, and external factors. We analyzed the factors influencing dimer-monomer equilibrium as well as the rate of interconversion, using the scFv[McPC603 as a model system. In this molecule, the stability of the VH-VL interaction can be conveniently varied by adjusting the ionic strength (because of its influence on the hydrophobic effect), by pH (presumably because of the presence of titratable groups in the interface), and by the presence or absence of the antigen phosphorylcholine, which can be rapidly removed due to its very fast off-rate. It was found that the monomer is the thermodynamically stable form with linkers of 15 and 25 amino acids-length under all conditions tested (35 &mgr;M or less). The dimer is initially formed in periplasmic expression, presumably by domain swapping, and can be trapped by all factors which stabilize the VH-VL interface, such as the presence of the antigen, high ionic strength, and pH below 7.5. Under all other conditions, it converts to the monomer. Predominantly monomer is obtained during in vitro folding. Monomer is stabilized against dimerization at very high concentrations by the same factors which stabilize the VH-VL interaction. These results should be helpful in producing molecules with defined oligomerization states.

L9 ANSWER 9 OF 20 MEDLINE DUPLICATE 5

ACCESSION NUMBER: 1998443242 MEDLINE
DOCUMENT NUMBER: 98443242 PubMed ID: 9769213
TITLE: Reproducing the natural evolution of protein structural features with the selectively infective phage (SIP) technology. The kink in the first strand of antibody kappa domains.
AUTHOR: Spada S; Honegger A; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut der Universität Zurich,
Winterthurerstrasse 190, Zurich, CH-8057, Switzerland.
SOURCE: JOURNAL OF MOLECULAR BIOLOGY, (1998 Oct 23) 283 (2)
395-407.
Journal code: J6V; 2985088R. ISSN: 0022-2836.
PUB. COUNTRY: ENGLAND; United Kingdom
Journal; Article; (JOURNAL ARTICLE)
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199811
ENTRY DATE: Entered STN: 19990106
Last Updated on STN: 19990106
Entered Medline: 19981125

AB The beta-sandwich structure of immunoglobulin variable domains is characterized by a typical kink in the first strand, which allows the first part of the strand to hydrogen bond to the outer beta-sheet (away from the VH-VL interface) and the second part to the inner beta-sheet. This kink differs in length and sequence between the V_{kappa}, V_{lambda} and VH domains and yet is involved in several almost perfectly conserved interactions with framework residues. We have used the selectively infective phage (SIP) system to select the optimal kink region from several defined libraries, using an anti-hemagglutinin single-chain Fv (scFv) fragment as a model system. Both for the kink with the V_{kappa} domain length and that with the V_{lambda} length, a sequence distribution was selected that coincides remarkably well with the sequence distribution of natural antibodies. The selected scFv fragments were purified and characterized, and thermodynamic stability was found to be the prime factor responsible for selection. These data show that the SIP technology can be used for optimizing protein structural features by evolutionary approaches.

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L9 ANSWER 10 OF 20 MEDLINE DUPLICATE 6

ACCESSION NUMBER: 1998437373 MEDLINE
DOCUMENT NUMBER: 98437373 PubMed ID: 9761676
TITLE: The nature of antibody heavy chain residue H6 strongly influences the stability of a VH domain lacking the disulfide bridge.
AUTHOR: Langedijk A C; Honegger A; Maat J; Planta R J;
van Schaik R C; Pluckthun A
CORPORATE SOURCE: Biochemisches Institut Universität Zurich,

SOURCE: Winterthurerstrasse 190, Baar, CH-8057, Switzerland.
JOURNAL OF MOLECULAR BIOLOGY, (1998) 283 (1) 95-110.
Journal code: JMB; 2985088R. ISSN: 0022-2836.
PUB. COUNTRY: ENGLAND: United Kingdom
LANGUAGE: English
FILE SEGMENT: Journal; Article; (JOURNAL ARTICLE)
ENTRY MONTH: Priority Journals
199811
ENTRY DATE: Entered STN: 19990106
Last Updated on STN: 19990106
Entered Medline: 19981105

AB Monoclonal antibody mAb 03/01/01, directed against the musk odorant traseolide, carries a serine residue instead of the conserved Cys H92 in the heavy chain variable domain, and is thus lacking the highly conserved disulfide bridge. We investigated the energetic consequence of restoring the disulfide bond and the nature of residue H6 (Glu or Gln), which is poised to interact with Ser H92 in the recombinant scFv fragment obtained from this antibody. In the scFv fragment derived from this antibody, the stabilizing effect of Gln H6 over Glu was found to be as large as the effect of reintroducing the disulfide bond. We have analyzed the conformation and hydrogen bond pattern of Gln H6 and Glu H6 in antibodies carrying these residues and suggest mechanisms by which this residue could contribute to VH domain stability. We also show that the unpaired cysteine H22 is buried, and conforms to the expected VH structure. The antibody appears to have acquired two somatic mutations (Ser H52 and Arg H66), which had been previously characterized as having a positive effect on VH stability. The overall domain stability is the decisive factor for generating functional, disulfide-free antibody domains, and several key residues play dominant roles.

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L9 ANSWER 11 OF 20 MEDLINE DUPLICATE 7
ACCESSION NUMBER: 1998161424 MEDLINE
DOCUMENT NUMBER: 98161424 PubMed ID: 9502319
TITLE: Parallel pathways in the folding of a short-term denatured scFv fragment of an antibody.
AUTHOR: Freund C; Gehrig P; Baici A; Holak T A; Pluckthun A
CORPORATE SOURCE: Department of Biochemistry, University of Zurich, Switzerland.
SOURCE: FOLDING AND DESIGN, (1997) 3 (1) 39-49.
Journal code: CUD; 9604387, ISSN: 1359-0278.
PUB. COUNTRY: ENGLAND: United Kingdom
LANGUAGE: English
FILE SEGMENT: Journal; Article; (JOURNAL ARTICLE)
ENTRY MONTH: Priority Journals
199804
ENTRY DATE: Entered STN: 19980416
Last Updated on STN: 19980416
Entered Medline: 19980408

AB BACKGROUND: Antibodies are prototypes of multimeric proteins and consist of structurally similar domains. The two variable domains of an antibody (VH and VL) interact through a large hydrophobic interface and can be expressed as covalently linked single-chain Fv (scFv) fragments. The in vitro folding of scFv fragments after long-term denaturation in guanidinium chloride is known to be slow. In order to delineate the nature of the rate-limiting step, the folding of the scFv fragment of an antibody after short-term denaturation has been investigated. RESULTS: Secondary structure formation, measured by H/D-exchange protection, of a mutant scFv fragment of an antibody after short incubation in 6 M guanidinium chloride was shown to be multiphasic. NMR analysis shows that an intermediate with significant proton protection is observed within the dead time of the manual mixing experiments. Subsequently, the folding reaction proceeds via a biphasic reaction and mass spectrometry analyses of the exchange experiments confirm the existence of two parallel pathways. In the presence of cyclophilin, however, the faster of the two phases vanishes (when followed by intrinsic tryptophan fluorescence), while the slower phase is not significantly enhanced by equimolar cyclophilin. CONCLUSIONS: The formation of an early intermediate, which shows amide-proton exchange protection, is independent of proline isomerization. Subsequently, a proline cis-trans isomerization reaction in the rapidly formed intermediate, producing 'non-native' isomers, competes with the fast formation of native species. Interface formation in a folding intermediate of the scFv fragment is proposed to prevent the back-isomerization of these prolines from being efficiently catalyzed by cyclophilin.

L9 ANSWER 12 OF 20 MEDLINE DUPLICATE 8
ACCESSION NUMBER: 97337429 MEDLINE
DOCUMENT NUMBER: 97337429 PubMed ID: 9194169
TITLE: Disrupting the hydrophobic patches at the antibody variable/constant domain interface: improved in vivo folding and physical characterization of an engineered scFv fragment.
AUTHOR: Nieba L; Honegger A; Krebber C;
Pluckthun A
CORPORATE SOURCE: Biochemisches Institut, Universität Zurich, Switzerland.
SOURCE: PROTEIN ENGINEERING, (1997 Apr) 10 (4) 435-44.
Journal code: PRI; 8801484. ISSN: 0269-2139.
PUB. COUNTRY: ENGLAND: United Kingdom
LANGUAGE: English
FILE SEGMENT: Journal; Article; (JOURNAL ARTICLE)
ENTRY MONTH: Priority Journals
199708
ENTRY DATE: Entered STN: 19970902
Last Updated on STN: 19970902
Entered Medline: 19970818

AB By constructing Fv and single-chain Fv (scFv) fragments of antibodies, the variable domains are taken out of their natural context in the Fab fragment, where they are associated with the constant domains of the light (CL) and heavy chain (CH1). As a consequence, all residues of the former variable/constant domain interface become solvent exposed. In an analysis of 30 non-redundant Fab structures it was found that at the former variable/constant domain interface of the Fv fragment the frequency of exposed hydrophobic residues is much higher than in the rest of the Fv fragment surface. We investigated the importance of these residues for different properties such as folding in vivo and in vitro, thermodynamic stability, solubility of the native protein and antigen affinity. The experimental model system was the scFv fragment of the anti-fluorescein antibody 4-4-20, of which only 2% is native when expressed in the periplasm of Escherichia coli. To improve its in vivo folding, a mutagenesis study of three newly exposed interfacial residues in various combinations was carried out. The

SOURCE: JOURNAL OF MOLECULAR BIOLOGY (1994 Sep 16) 242 (2) 165-74.
Journal code: J6V; 2985088R. ISSN: 0022-2836.
PUB. COUNTRY: ENGLAND: United Kingdom
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199410
ENTRY DATE: Entered STN: 19941031
Last Updated on STN: 19970203
Entered Medline: 19941014

AB By thermal equilibrium measurements we found a three-state folding behavior of mature Escherichia coli beta-lactamase TEM2. The thermodynamically stable intermediate H had no enzymatic activity, but a native-like secondary structure. State H was 9 kcal mol⁻¹ less stable than the native state N and 4 kcal mol⁻¹ more stable than the totally unfolded state U, which is consistent with urea equilibrium measurements of mature beta-lactamase measured under similar conditions. Between 38 degrees C and 50 degrees C there was a decrease in the apparent equilibrium constant for dissociation K'D of the complex between GroEL and mature beta-lactamase, at least partially caused by a decrease in the thermodynamic stability of the native form of mature beta-lactamase. GroEL-bound beta-lactamase was released either after addition of ATP, or in the presence of a competing substrate (i.e. a single-chain antibody), or after lowering the temperature. Whereas at 10 degrees C the folding reaction of mature beta-lactamase was rate limiting, at 37 degrees C the release reaction was the rate-determining step for the regain of beta-lactamase activity, consistent with a decrease of the equilibrium constant for dissociation KD of the complex with temperature. A temperature dependent behavior of GroEL was also observed, when measuring the anilinonaphthalene sulfonic acid (ANS) fluorescence of the chaperone. Similar to all other substrate proteins studied so far, the maximal tryptophan fluorescence of GroEL-bound beta-lactamase was observed at 342 nm. Our results are compatible with a hydrophobic binding pocket of GroEL and confirm the suggested thermodynamic partitioning model for hydrophobic binding of polypeptides by GroEL.

L9 ANSWER 17 OF 20 MEDLINE
ACCESSION NUMBER: 92292158 MEDLINE
DOCUMENT NUMBER: 92292158 PubMed ID: 1602480
TITLE: Refined crystal structure of a recombinant immunoglobulin domain and a complementarity-determining region 1-grafted mutant.
AUTHOR: Steipe B; Pluckthun A; Huber R
CORPORATE SOURCE: Abteilung Strukturforschung, Max-Planck-Institut für Biochemie, Martinsried, Germany.
SOURCE: JOURNAL OF MOLECULAR BIOLOGY, (1992 Jun 5) 225 (3) 739-53.
Journal code: J6V; 2985088R. ISSN: 0022-2836.
PUB. COUNTRY: ENGLAND: United Kingdom
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199207
ENTRY DATE: Entered STN: 19920724
Last Updated on STN: 19980206
Entered Medline: 19920710

AB We report the solution of the crystal structure of a mutant of the immunoglobulin VL domain of the antibody McPC603, in which the complementarity-determining region 1 segment is replaced with that of a different antibody. The wild-type and mutant crystal structures have been refined to a crystallographic R-factor of 14.9% at a nominal resolution of 1.97 Å. A detailed description of the structures is given. Crystal packing results in a dimeric association of domains in a fashion closely resembling that of an Fv fragment. The comparison of this VL domain with the same domain in the Fab fragment of McPC603 shows that the structure of an immunoglobulin VL domain is largely independent of its mode of association, even in places where the inter-subunit contacts are not conserved between VL and VH. In all three complementarity-determining regions we observe conformations that would not have been predicted by the canonical structure hypothesis. Significant differences between the VL domain dimer and the Fab fragment in the third complementarity-determining region show that knowledge of the structure of the dimerization partner and its exact mode of association may be needed to predict the precise conformation of antigen-binding loops.

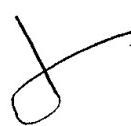
L9 ANSWER 18 OF 20 MEDLINE DUPLICATE 12
ACCESSION NUMBER: 91175756 MEDLINE
DOCUMENT NUMBER: 91175756 PubMed ID: 1901023
TITLE: Mapping and modification of an antibody hapten binding site: a site-directed mutagenesis study of McPC603.
AUTHOR: Glockshuber R; Stadlmüller J; Pluckthun A
CORPORATE SOURCE: Gzenzentrum der Universität München, Max-Planck-Institut für Biochemie, Martinsried, West Germany.
SOURCE: BIOCHEMISTRY, (1991 Mar 26) 30 (12) 3049-54.
Journal code: AOG; 0370623. ISSN: 0006-2960.
PUB. COUNTRY: United States
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199105
ENTRY DATE: Entered STN: 19910519
Last Updated on STN: 19910519
Entered Medline: 19910501

AB The quantitative contributions of various amino acid residues to hapten binding in the Fv fragment of the antibody McPC603 were investigated by site-directed mutagenesis. The three-dimensional structure of the Fab' fragment of McPC603 is known to atomic resolution. The haptens phosphocholine, choline sulfate, 3-(trimethylammonium)propane-1-sulfonate, 4-(trimethylammonium)butyric acid, and 4-(trimethyl-ammonium)butyric acid methyl ester were tested for binding. It was found that the phosphate group but not the sulfate and sulfonate groups, interacts with the hydroxyl group of Tyr33(h). The required positive charge for the binding of the phosphate must be contributed by Arg52(h); a lysine at this position or an additional positive charge at position 33(h) abolishes the binding to a phosphocholine affinity column. The interaction between Tyr100(l) and Glu35(h) was found to be essential and could not be functionally replaced by any other pair of residues tested. Binding of the quaternary ammonium ion needs a negative charge; it can reside in either Asp97(l) or Asp101(h), but both together prevent binding to the affinity column. These data may serve as the basis for the development of quantitative treatments of antigen-antibody interactions.

L9 ANSWER 19 OF 20 MEDLINE
ACCESSION NUMBER: 92070228 MEDLINE

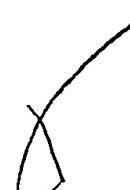
DOCUMENT NUMBER: 92070228 PubMed ID: 195
 TITLE: Catalytic antibodies: contributions from engineering and expression in *Escherichia coli*.
 AUTHOR: Pluckthun A; Stadtmuller J
 CORPORATE SOURCE: Genzentrum, Universitat Munchen, Max-Planck-Institut fur Biochemie, Martinsried, Federal Republic of Germany.
 SOURCE: CIBA FOUNDATION SYMPOSIUM, (1991) 159 103-12; discussion 112-7. Ref: 51
 PUB. COUNTRY: Netherlands
 Journal; Article; (JOURNAL ARTICLE)
 General Review; (REVIEW)
 (REVIEW, TUTORIAL)
 LANGUAGE: English
 FILE SEGMENT: Priority Journals
 ENTRY MONTH: 199201
 ENTRY DATE: Entered STN: 19920124
 Last Updated on STN: 19920124
 Entered Medline: 19920109

AB Antibodies have been raised against the transition state of many reactions and shown to catalyse the relevant reaction. Their moderate catalytic efficiencies can be increased by protein engineering, if ways can be found to express the engineered antibody. We have developed a system by which fully functional Fv and Fab fragments can be expressed in *Escherichia coli*. The Fv fragment dissociates at low concentrations; we therefore devised methods to stabilize the fragment. We showed that the Fv fragment of the antibody McPC603, a phosphorylcholine-binding immunoglobulin A, binds the antigen with the same affinity as does the intact antibody isolated from mouse ascites. Phosphorylcholine is an analogue of the transition state for the hydrolysis of choline carboxylate ester. The Fv fragment of McPC603 catalysed this hydrolysis. Mutational analysis of the residues in the binding site of the antibody has shown which are essential for binding and for catalysis, and the importance of charged residues in certain positions. The *E. coli* expression system combined with protein engineering and screening methods will facilitate understanding of enzyme catalysis and the development of new catalytic antibodies



L9 ANSWER 20 OF 20 MEDLINE
 ACCESSION NUMBER: 91264723 MEDLINE
 DOCUMENT NUMBER: 91264723 PubMed ID: 2096820
 TITLE: Properties of Fv and Fab fragments of the antibody McPC603 expressed in *E. coli*.
 AUTHOR: Pluckthun A; Glockshuber R; Skerra A; Stadtmuller J
 CORPORATE SOURCE: Genzentrum der Universitat Munchen, Germany.
 SOURCE: BEHRING INSTITUTE MITTEILUNGEN, (1990 Dec) (87) 48-55.
 PUB. COUNTRY: GERMANY: Germany, Federal Republic of
 Journal; Article; (JOURNAL ARTICLE)
 LANGUAGE: English
 FILE SEGMENT: Priority Journals
 ENTRY MONTH: 199107
 ENTRY DATE: Entered STN: 19910802
 Last Updated on STN: 19970203
 Entered Medline: 19910717

AB The Fv and Fab fragments of the phosphorylcholine binding antibody McPC603 were functionally expressed in *E. coli*. This was achieved by the co-expression and co-secretion of both chains to the periplasm, where correct processing, folding and assembly occurred. Interestingly, the fraction of correctly folded Fab fragment is smaller than that of the Fv fragment in *E. coli*. The intrinsic hapten binding affinity was shown to be identical for the recombinant Fv or Fab fragment, the whole antibody and the Fab fragment obtained by proteolysis from the mouse antibody. Fluorescence and crosslinking analyses showed that the Fv fragment dissociates at high dilution, but that it is stabilized by hapten binding. The recombinant Fv fragment was shown to have catalytic activity to hydrolyze choline-p-nitrophenyl carbonate and constitutes therefore a promising model system with which the structural requirements of catalytic antibodies can be studied by altering the protein itself.



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(FILE 'HOME' ENTERED AT 07:17:38 ON 11 JUN 2001)

FILE 'MEDLINE, CAPLUS, EMBASE, BIOSIS' ENTERED AT 07:17:58 ON 11 JUN 2001
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 L2 47308 S L1 (P) (DOMAIN OR FRAGMENT)
 L3 7807 S L2 (P) DNA
 L4 17 S L3 (P) INTERFACE
 L5 6 DUP REM L4 (11 DUPLICATES REMOVED)
 L6 651 S PLUCKTHUN A?/AU OR NIEBA L?/AU OR HONEGGER A?/AU
 L7 313 S L6 AND ANTIBOD?
 L8 46 S L7 AND HYDRO?
 L9 20 DUP REM L8 (26 DUPLICATES REMOVED)

=> logoff

ALL L# QUERIES AND ANSWER SETS ARE DELETED AT LOGOFF

LOGOFF? (Y)/N/HOLD:y

COST IN U.S. DOLLARS	SINCE FILE ENTRY	TOTAL SESSION
FULL ESTIMATED COST	48.54	48.69
DISCOUNT AMOUNTS (FOR QUALIFYING ACCOUNTS)	SINCE FILE ENTRY	TOTAL SESSION
CA SUBSCRIBER PRICE	-1.76	-1.76

STN INTERNATIONAL LOGOFF AT 07:27:21 ON 11 JUN 2001